



<https://doi.org/10.15407/ukrbotj83.03.161>

RESEARCH ARTICLE

AFLP-based genetic diversity and population structure of arctic-alpine relict species *Dryas octopetala* (Rosaceae) in Ukraine

Andrii TARIEIEV^{1,2} , Roman CHEREPANYN^{3,4} 

¹ Department of Forest Genetics and Forest Tree Breeding, University of Göttingen,
2 Büsgenweg, Göttingen 37077, Germany

² Institute of Biology, Geobotany and Botanical Garden, Martin Luther University Halle-Wittenberg,
1 Am Kirchtor, Halle (Saale) 06108, Germany

³ International Research and Educational Centre “Observatory”,
Biology and Ecology Department, Vasyl Stefanyk Carpathian National University,
57 Shevchenko Str., Ivano-Frankivsk 76018, Ukraine

⁴ WWF-Ukraine (Public Union World Wide Fund for Nature Ukraine),
4 Raisy Okipnoi Str., office 170, Kyiv 02002, Ukraine

Addresses for correspondence: AT, andrii.tarieiev@botanik.uni-halle.de; RC, roman.cherepanyn@cnu.edu.ua

Abstract. Arctic-alpine plant communities host many endemic and relict species. Many such species are sensitive to habitat changes and have been already affected by climate change, which makes their study especially relevant. The current study focuses on the circumpolar arctic-alpine relict diploid subshrub / dwarf shrub *Dryas octopetala* (Rosaceae) in Ukraine. Four known and currently available populations were assessed and sampled. Three sets of 50 different AFLP markers each were used to investigate genetic diversity, population structure, and other parameters. STRUCTURE analyses, in conjunction with the phylogenetic network, indicate no significant differences in genetic structure among populations. However, the population on Mt. Brebeneskul shows a very marginal difference in comparison to three other dryad populations in Ukraine. Geographic isolation, differing migration paths, and the potential for interbreeding could explain this outcome. Additional studies using various methods (e.g., SSR, genome-wide SNP analysis, GWAS, WGS) are needed to obtain a clearer picture.

Keywords: AFLP, Carpathians, *Dryas octopetala*, STRUCTURE, PCoA, populations, Ukraine

Introduction

Arctic-alpine plants play a crucial role in the ecosystems of the Arctic and subarctic regions, as well as the mountains of the temperate zone of the

Northern Hemisphere. The term “arctic-alpine species” reflects a geographic distribution and encompasses a range of species, from widespread, community-forming species to rare, endemic, and relict ones. Some could be used as model organisms in

ARTICLE HISTORY. Submitted 22 February 2026. Revised 26 May 2026. Published 24 June 2026

CITATION. Tarieiev A., Cherepanyn R. 2026. AFLP-based genetic diversity and population structure of arctic-alpine relict species *Dryas octopetala* (Rosaceae) in Ukraine. *Ukrainian Botanical Journal*, 83(3): 161–171. <https://doi.org/10.15407/ukrbotj83.03.161>

© M.G. Kholodny Institute of Botany, NAS of Ukraine, 2026

© Publisher PH "Akademperiodyka" of the NAS of Ukraine, 2026

This is an open access article under the CC BY license (<https://creativecommons.org/licenses/by/4.0/>)

studies on ecosystem responses to climate change, which is especially relevant today. Climate change and high anthropogenic pressure often reduce the distribution areas and alter the population structures of rare arctic-alpine species (Cherepanyn, 2017; Kozłowski, 2024).

Dryas octopetala L. (*Rosaceae*) is a circumpolar (or mainly Eurasian, in the strict sense) arctic-alpine relict diploid dwarf shrub that occurs in the Arctic and Subarctic zones and the mountains of the Northern Hemisphere in the alpine and subalpine zones of Eurasia (replaced by related species in continental North America: see Springer, Parfitt, 2014), mainly on limestone and sandstone rocks with carbonate content, stony slopes with weak soil, and a microphytoclimate present only in the surface horizon (McGraw, Antonovics, 1983; McGraw, 1987). It is an insect-pollinated mesoxerophyte and hemiphanerophyte with seed dispersal by wind (anemochory). In Ukraine, the mountain avens (*Dryas*) are considered rare and are protected at the national level. The species is listed in the *Red Data Book of Ukraine* (Andrienko, Mosyakin, 2009). The *Dryadeta octopetalae* plant community is also protected at the national level and listed in the *Green Data Book of Ukraine* (2009).

Populations of *Dryas octopetala* occur on high mountain meadows and often are dominant in *Achilleo Schurii-Dryadetum* plant communities (*Carrici rupestris-Kobresietea ballardii* class) and *Loiseleurio-Vaccinietaea* class communities. The species usually occurs together with *Carex sempervirens* Vill., *Campanula alpina* Jacq., *Vaccinium myrtillosum* L., *V. vitis-idea* L., *Anemonastrum narcissiflorum* (L.) Holub (= *Anemone narcissiflora* L.), *Bartsia alpina* L., *Homogyne alpina* (L.) Cass., *Sesleria coerulans* Friv., *Salix herbacea* L., etc. It often forms cushion turfs, and the spatial distribution of individuals in populations is uneven (Chopyk, 1976).

Populations of *D. octopetala* have two strategies of propagation, vegetative and generative ones. The first one is crucial for the self-maintenance of populations and leads to the formation of dense clones, genetically identical plants that could even be considered a single individual or super-individual. Genets (vegetative clones) can remain stable and alive for a long time, with some estimates suggesting a lifespan of over 500 years (de Witte et al., 2012). This propagation is typical for many highland and montane species (Tsaryk et al., 2004). Generative propagation is less common, but it is crucial for

maintaining genetic diversity and dispersal. It is also reported that *D. octopetala* has high clonal diversity (de Witte et al., 2021)

Historic occurrence of *D. octopetala* in Ukraine, even during the Allerød–Holocene, was much wider than its current distribution, as demonstrated by generalized data of paleopalynofloristic studies (Bezusko et al., 2011, 2025). At present, there are four known localities of *D. octopetala* in the Ukrainian Carpathians, which are located within two mountain ranges: the Svydovets (Blyznytysia and Zhandarmy mountains) and Chornohora (Brebeneskul [Brebenescul in Romanian] and Pip Ivan mountains). These populations occur at high elevations (1645–1988 m a.s.l.) and are associated with alpine and subalpine habitats characterized by rocky substrates and open vegetation.

Previous studies have shown that the population structure and regeneration of *D. octopetala* are influenced by habitat conditions, particularly the availability of open microsites and the degree of competition with the dense grass cover (Cherepanyn, 2018, 2019). In general, exposed and sparsely vegetated areas promote both vegetative and generative reproduction, whereas dense turf-forming species may limit seedling establishment.

In a significant portion of the arctic-alpine and relict species at the southern limit of their distribution ranges in the mountains of temperate latitudes, there is a narrowing or shrinking of the population ranges, changes in the population structure and dynamics due to global warming (Pauli et al., 2007; Erschbamer et al., 2009). The ability to disperse over long distances and to occupy certain ecological niches in mainly unsuitable habitats is crucial for arctic-alpine species for their survival under climate change (Marcysiak, 2010). Due to climate change, species' population structures and vegetation cover change with increasing the elevation above sea level. There is an increase in the occurrence of the shrubs *Pinus mugo* Turra, *Alnus alnobetula* (Ehrh.) K. Koch (= *Alnus viridis* (Chaix.) DC.), and *Juniperus communis* L. var. *saxatilis* Pall. at the border of the subalpine and alpine zones. Additionally, declines in population vitality and shifts in the population structure of some high-altitude species in high mountain meadows have been observed (Kobiv, 2017, 2018; Cherepanyn, 2018, 2019; Kyyak, Shtupun, 2021). Monitoring data within the framework of the GLORIA project (GLobal Observation Research Initiative in Alpine Environments)

indicate that the habitat areas of populations of alpine, nival, and subnival species are decreasing (Harald et al., 2007).

It is essential to understand the genetic structure of these species for developing efficient measures for protecting their populations. Such information is crucial when analyzing small populations, where gene drift often occurs, leading to a significant decrease in genetic diversity (Avise, 2008). Additionally, genetic research can provide insights into the mechanisms of gene information exchange, evolutionary and geographical processes within a species, and prospects for the survival of populations (Kreuzer, 2014). Therefore, further molecular genetic research is needed to analyze such issues.

Today, molecular ecological research is particularly popular in Europe in the context of studying rare, endemic, relict, and peripheral-range (marginal) plant species, including *D. octopetala* (De Witte et al., 2012; Varsamis et al., 2021). AFLP data consist of anonymous multilocus dominant marker sets, scored as presence-absence (Vos et al., 1995; Kück et al., 2012). They are commonly used in evolutionary, ecological, and population genetic studies (Mueller, Wolfenbarger, 1999; Meudt, Clarke, 2007). Unfortunately, the studies of the genetic structure of populations in high-altitude species still remain insufficiently represented in Ukraine. This work aims to address the issue of the molecular genetic diversity of small populations of rare plant species in the Ukrainian Carpathians, particularly the high-altitude dwarf shrub *D. octopetala*.

Materials and Methods

Collection of samples. Sampling in *Dryas octopetala* habitats was conducted in 2018 during the growing season, mainly in July and August, across the whole distribution area of *D. octopetala* in Ukraine, namely from 4 populations of the species in the Ukrainian Carpathians (Table 1, Fig. 1). Populations were defined as groups of plants separated within one or different mountain massifs. Populations were sampled evenly along the transects to cover the entire habitat area of the species. To achieve statistically representative sampling, 30 samples were collected per population within one growing season (120 samples from 4 populations). One or two well-developed leaf blades without any visual signs of damage, fungal infestation, or necroses were carefully collected from plants.

Then, the material from each individual was immediately placed in separate bags with silica gel for rapid drying. At the next step, bags with collected samples from each population were grouped and stored in sealed plastic bags with silica gel (Chase, Hills, 1991). The packed material was stored at room temperature (with periodic replacement of the silica gel if necessary).

AFLP-analysis. In 2019, total genomic DNA was extracted from tissue powder using the Qiagen DNeasy Plant Mini Kit (Qiagen, Germany <http://www.qiagen.com/>) according to the manufacturer's protocol.

For the subsequent molecular genetic analysis of the populations, the analysis based on amplified fragment length polymorphism (AFLP) was performed according to the protocol/procedure developed by Vos et al. (1995) and described in detail in Cieślak et al. (2007, 2015), Meudt and Clarke (2007), Sucher et al. (2012), Paun and Schönswetter (2012). The following markers were selected for AFLP analysis of *D. octopetala*: ATG_CTG_K1, ATC_CAA_K2, ATC_CTG_K3. Fragment analysis was performed with capillary electrophoresis on ABI XL3500 (Applied Biosystems, USA).

Data analysis. AFLP profiles were analyzed using Genographer 2.1 software (Benham, 2001), which read DNA profiles in the 50–500 bp range and coded the presence/absence of profile peaks into a binary matrix of feature absence/presence (0/1).

The obtained binary matrices (Supplementary Material, Matrices S1–S4) were analyzed separately and then combined. Clustering analysis (Neighbour-Joining, with 1000 bootstrap iterations) was performed in PAST 4.16 (Hammer et al., 2001; Hammer, Harper, 2024). Principal coordinate analysis (PCoA) — in GenAlEx 6.51b2 (Peakall, Smouse 2006, 2012). Estimation of genetic structure and inference of basic statistics were performed using AFLP-SURV 1.0. (Vekemans, 2002; Vekemans et al., 2002; <https://ebe.ulb.ac.be/ebe/AFLP-SURV.html>). Population analysis was performed in STRUCTURE 2.3.4. (Porrás-Hurtado et al., 2013; Novembre, 2016; <https://web.stanford.edu/group/pritchardlab/structure.html>) on the GWGD HPC cluster using the StrAuto 1.0 python utility to automate and parallelize the STRUCTURE analysis (Chhatre, Emerson, 2017; <https://vc.popgen.org/software/strauto/>) with the following parameters: maxpops = 5, mcmc = 500000, burnin = 100000,

Table 1. **Physical and geographical characteristics of *Dryas octopetala* locations**
 (*determined using a GPS navigator “Garmin eTrex 30x”)

Population locality	Height above sea level, m	Exposure of the slope	Area of the habitats, m ²	Latitude*	Longitude*
Brebeneskul (Chornohora mountain range)	1941	north-west	250	N48°06'17.1"	E024°34'01.7"
Pip Ivan (Chornohora mountain range)	1988	south-west	150	E024°37'36.6"	
Blyznytsia (Svydovets mountain range)	1803	north-east	350	N48°13'09.7"	E024°13'53.3"
Zhandarmy rocks (Svydovets mountain range)	1645	south-east	1250	N48°13'44.0"	E024°13'53.3"

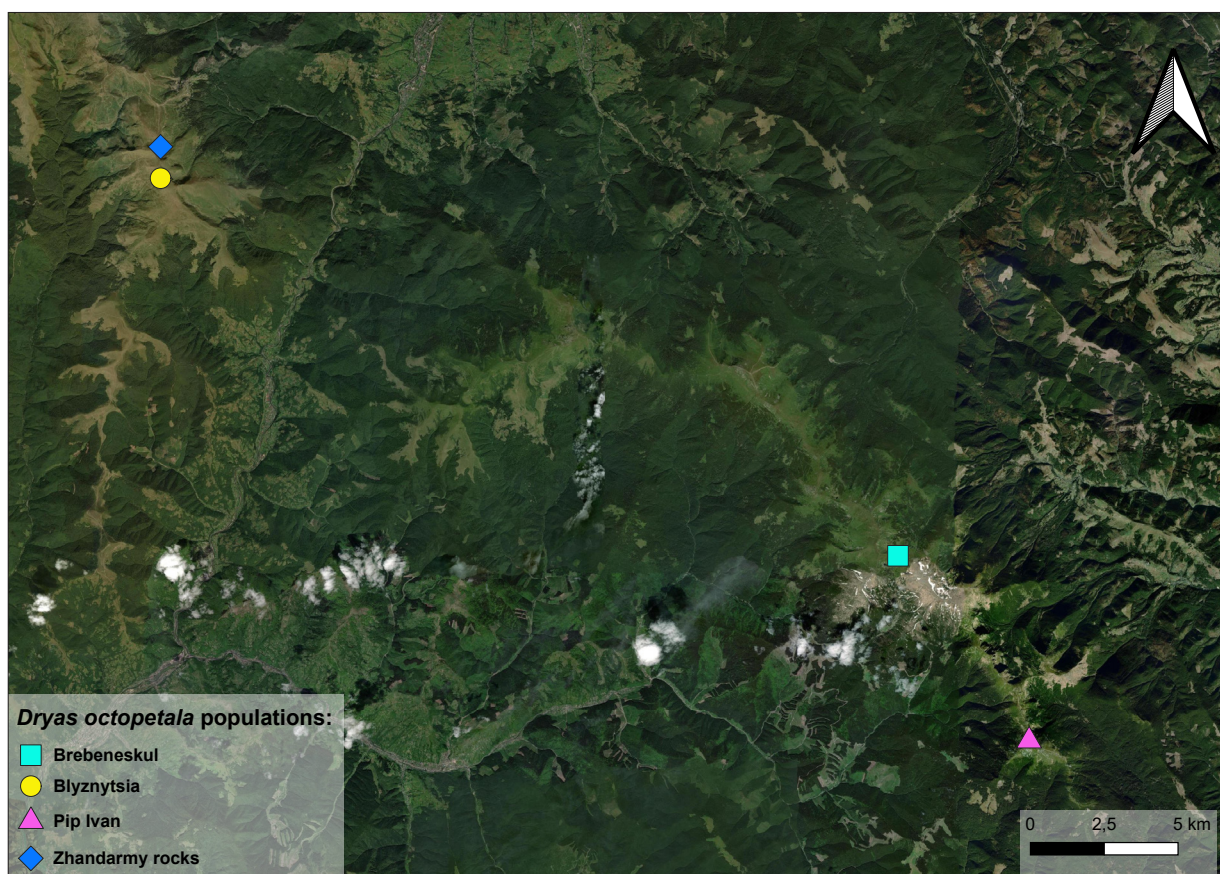


Fig. 1. Locations of the populations of *Dryas octopetala* (created in QGIS 3.44, “Solothurn” based on ESRI Satellite map)

K-runs = 10. STRUCTURE output was analyzed and visualised using the Clustering Markov Package Across K (CLUMPAK, Kopelman et al., 2015) server (<https://tau.evolveq.net/clumpak/>). Identification of prospective best K values was performed in StructureSelector (Li, Liu, 2018; [\[ac.cn/StructureSelector/\]\(https://lmme.ac.cn/StructureSelector/\)\) based on several different approaches: Evanno \$\Delta K\$ method \(Evanno et al., 2005\), Choose K method \(Raj et al., 2014\), and Puechmaille method with estimation of four different parameters — medMedK, maxMedK, med-MeanK, and maxMeanK \(Puechmaille, 2016\).](https://lmme.</p>
</div>
<div data-bbox=)

In addition, we converted binary matrices to nexus format and reconstructed the following types of haplotype networks using PopART 1.7 (<https://popart.maths.otago.ac.nz/>, Leigh, Bryant, 2015): Minimum Spanning Network, Median Joining Network (Bandelt et al., 1999), Integer NJ Net (French et al., 2014; Leigh, Bryant, 2015), and TCS (Clement et al., 2000, 2002). Network types were inferred for each marker set separately and for the combined set.

Results

Cluster analysis. Pairwise genetic distances (Nei's genetic distance after Lynch and Milligan (1994); and Reynolds, Weir, and Cockerham's (1983) genetic distance, respectively, Table 2) were the smallest between *Dryas octopetala* populations on Blyznytsia and Pip Ivan mountains (0.0081 and 0.0326), while the largest were between populations on Pip Ivan

Table 2. **Genetic distances between populations of *Dryas octopetala***

(bold — Nei's genetic distance after Lynch and Milligan (1994); italics — Reynolds, Weir, and Cockerham's genetic distance)

<i>Dryas octopetala</i> populations	Brebeneskul (Chornohora mountain range)	Pip Ivan (Chornohora mountain range)	Blyznytsia (Svydovets mountain range)	Zhandarmy rocks (Svydovets mountain range)
Brebeneskul (Chornohora mountain range)		<i>0.1077</i>	<i>0.0717</i>	<i>0.1144</i>
Pip Ivan (Chornohora mountain range)	0.0268		<i>0.0326</i>	<i>0.1352</i>
Blyznytsia (Svydovets mountain range)	0.0182	0.0081		<i>0.1200</i>
Zhandarmy rocks (Svydovets mountain range)	0.0259	0.0313	0.0286	

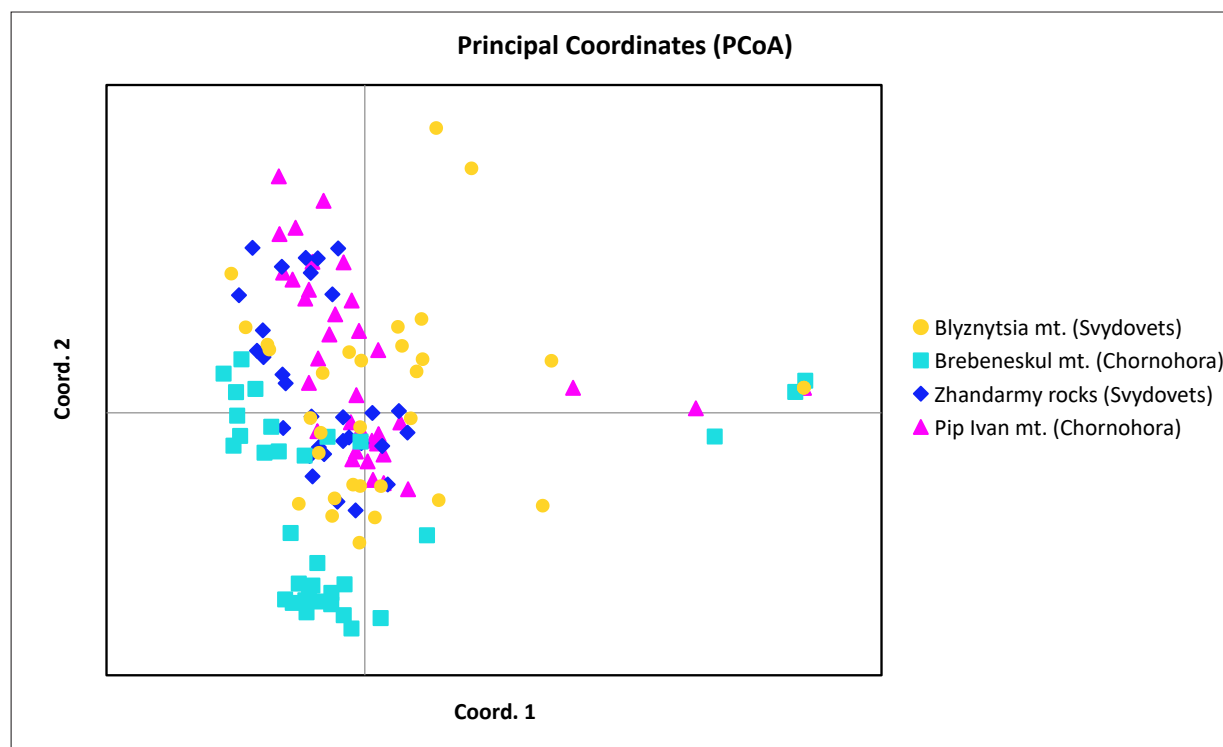


Fig. 2. Principal Coordinate Analysis (PCoA) based on a combined set of markers for all populations of *Dryas octopetala*

and Zhandarmy rocks (0.0313 and 0.1352, respectively).

Pairwise F_{ST} values for all markers show little to moderate differentiation between populations and follow the same pattern as genetic distances, with the lowest differentiation between *D. octopetala* populations on Blyznytsia and Pip Ivan mountains (0.0321), and the highest between populations on Pip Ivan and Zhandarmy rocks (0.1264). F_{ST} values for the rest of the cases in ascending order: between populations on Blyznytsia and Brebeneskul — 0.0692; Pip Ivan and Brebeneskul — 0.1021; Brebeneskul and Zhandarmy rocks — 0.1081; Blyznytsia and Zhandarmy rocks — 0.1131.

Principal Coordinate Analysis (PCoA). PCoA based on a combined set of markers (Fig. 2) shows a distinct grouping of samples by population assignments. However, these clusters are not separated, and there is no geographic pattern (by mountain range) either.

STRUCTURE analysis. Different approaches to optimal K estimation produced different results (Table 3).

STRUCTURE analysis reveals differences between the analyzed populations (Fig. 3). For the best $K = 2$, the population on the Zhandarmy rocks (3) is entirely monomorphic. At the same time, the other three demonstrate some degree of heterogeneity. We were unable to identify any specific geographic pattern among the individuals assigned to the orange cluster.

In the case of $K = 3$ (optimal for Puchmaille's medMedK and medMeanK), the population on Zhandarmy rocks revealed two clusters, while the rest revealed three ones. As in the previous case, no specific geographic pattern was identified.

We did not identify any specific structure for $K = 4$ (optimal for Puchmaille's maxMedK and maxMeanK), which also corresponds to the number of actual populations.

In contrast, in the case of $K = 5$ (optimal for the Choose K method and optimal for Puchmaille's medMedK and medMeanK), the population on Mt. Brebeneskul (Chornohora mountain range) differs from all others: the majority of samples within it formed a separate cluster, which was almost absent in other populations. The remaining populations were similar, with only minor differences.

STRUCTURE analysis performed for separate populations and three separate sets of markers (50 loci each) presented in Supplementary Material (Figs. S1–S4), and generally agrees with the combined one.

In general, STRUCTURE analysis shows specific patterns of admixture and likely higher admixture rates within populations than between them (which is expected due to geographical isolation).

Haplotype networks. There is no apparent clustering in all the types of haplotype networks (TCS, NJ-Net, Minimum Spanning Network, Median Joining Network). Nevertheless, a specific population grouping pattern exists (Suppl. files S5–S8). At the same time, networks reveal significant differences in the number of reticulations across methods. In particular, NJ-Net (Suppl. file S6) is significantly more reticulated than the other types.

Discussion

The AFLP analysis of four different populations of *Dryas octopetala* in Ukraine revealed genetic differences within and among populations. However, neither a clear genetic structure nor significant genetic differences were detected. The results of PCoA, STRUCTURE analysis and different types of haplotype networks are generally in agreement and demonstrate similar patterns. There were minor differences in the STRUCTURE analysis outcomes depending on the optimal K. At $K = 2$, the population on the Zhandarmy rocks was monomorphic, whereas the rest were not. We assume the variation is random, as we could not identify any specific geographic pattern in the three populations (Blyznytsia, Pip Ivan, and Brebeneskul). At $K = 5$, the population on Brebeneskul Mt was distinct from the rest. Besides potential computational issues, such an outcome could be caused by geographic isolation, different migration paths, interbreeding, and/or other factors. To investigate further the possible admixture and other aspects, different markers and approaches are needed (in particular, genomic data could be helpful).

Since *Dryas octopetala* is protected and has a scattered distribution in Ukraine, genetic and genomic data could be useful for organizing specific protection/conservation measures. One key element for the long-term conservation of the species is to save and preserve as much genetic diversity as possible (Centre for Plant Conservation, 2019).

Additional studies, employing more powerful modern methods, such as high throughput whole-genome sequencing followed by different analyses (e.g., genome-wide SNP analyses, plastome & mitochondriome data, designing SSR panels, etc.)

Table 3. Optimal K estimation uses different methods for each population of *Dryas octopetala*, combining a set of markers

<i>Dryas octopetala</i> population marker sets	Evanno ΔK method	Choose the K method	Puchmaille method			
			medMedK	maxMedK	medMeanK	maxMeanK
Marker set 1, all populations	2	5	3–5	3–5	3–5	3–5
Marker set 2, all populations	2	4	1–3; 5	1–3; 5	1–3; 5	1–3; 5
Marker set 3, all populations	2	2	1–5	1–5	1–4	1–5
Population 1 (Blyznytisia), all markers	5	5	1–4	1–4	1–3	1–3
Population 2 (Brebneskul), all markers	2	3	1–4	1–4	1–3	1–3
Population 3 (Zhandarmy rocks), all markers	2	2	1–2	1–2	1–2	1–2
Population 4 (Pip Ivan), all markers	2	4	1–4	1–5	1–4	1–5
Combined, all populations and all markers	2	5	3; 5	4	3; 5	4

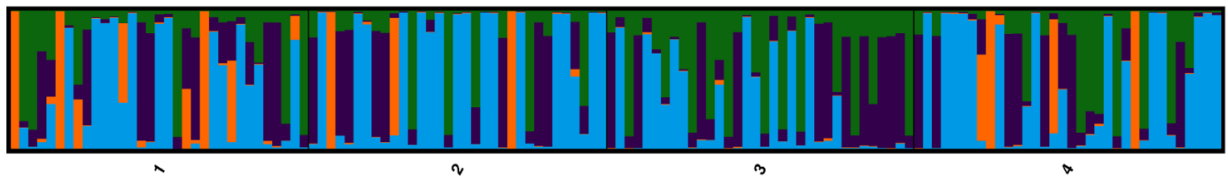
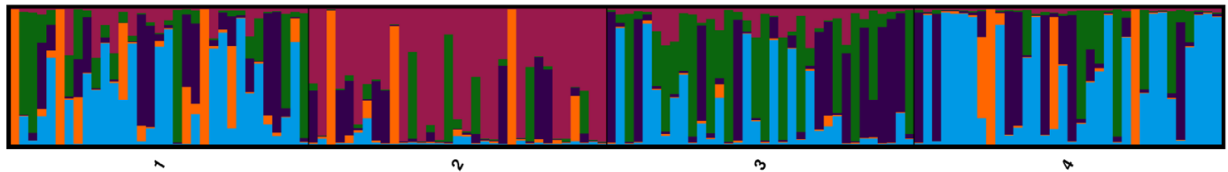
K=2**K=3****K=4****K=5**

Fig. 3. Genetic structure of populations of *Dryas octopetala* based on STRUCTURE analysis of combined data (150 loci, 4 populations). 1 — Blyznytisia Mt. (Svydovets mountain range); 2 — Brebneskul Mt. (Chornohora mountain range); 3 — Zhandarmy rocks (Svydovets mountain range); 4 — Pip Ivan Mt. (Chornohora mountain range)

and genome wide association studies (GWAS), are needed to obtain a clearer and more detailed picture. Currently, several genome assemblies are available for *Dryas*, including two for *D. octopetala* (White, Pirro, 2022; DToL), which are based on reference-grade whole-genome sequencing using PacBio long-read sequencing with 83x coverage (DToL; assemblies [GCA_963921425.1](https://ENA01.ebi.ac.uk/ena/record/GCA963921425.1) and [GCA_963921435.1](https://ENA01.ebi.ac.uk/ena/record/GCA963921435.1) in the European Nucleotide Archive, ENA). Modern long-read sequencing techniques not only make plant genome assemblies easier (Pucker et al., 2022), but they provide also epigenetic information as a byproduct which could be also useful for population studies (Liu, Zhong, 2024; Liu, Conesa, 2025). The availability of good reference genome assemblies makes it easier and cheaper to organize a separate study on the population genomics of mountain avens from Ukraine. In addition, such a project would be very relevant for several reasons: (1) there are no genomic studies of mountain avens in Ukraine and (2) the populations are relatively small, which makes them especially vulnerable. Although they are located in the Carpathians, there is a constant danger of destruction posed by the racist war of aggression against Ukraine.

Conclusions

AFLP analysis of four populations of *Dryas octopetala* in Ukraine reveals significant genetic similarity among them. No clear patterns were revealed within the populations. STRUCTURE analysis reveals specific differences depending on the optimal K value: with K = 2, the population on Zhandarmy rocks is entirely monomorphic, while under optimal K = 5, most samples from the population on Brebeneskul form a separate cluster. These results most probably indicate that disjunctions between the studied populations, with their genetic separation and isolation, occurred comparatively recently, probably already during some phases of the Holocene.

REFERENCES

- Andrienko T.L., Mosyakin S.L. 2009. *Dryas octopetala*. In: *Red Data Book of Ukraine. Plant Kingdom*. Ed. Ya.P. Didukh. Kyiv: Globalconsulting, p. 576. [Андрієнко Т.Л., Мосякін С.Л. 2009. *Dryas octopetala*. В кн.: Червона книга України. Рослинний світ. Ред. Я.П. Дідух. Київ: Глобалконсалтинг, с. 576.]
- Avise J.C. 2008. *Markery molekularne, historia naturalna i ewolucja*. Warszawa: WUW, 664 s. [Original edition: Avise J.C. 2004. *Molecular markers, natural history, and evolution*. 2nd ed. Sunderland (Massachusetts): Sinauer Associates Inc., xv + 684 pp.]
- Bandelt H., Forster P., Röhl A. 1999. Median-joining networks for inferring intraspecific phylogenies. *Molecular Biology and Evolution* 16(1): 37–48. <https://doi.org/10.1093/oxfordjournals.molbev.a026036>

Acknowledgements

Part of the work was conducted in 2019 within the framework of the Queen Jadwiga Foundation scholarship program at the Jagiellonian University, under the scientific supervision of Prof. Dr Hab. Józef Mitka (Institute of Botany, Jagiellonian University).

We express great gratitude to the "Molecular Biogeography and Systematics Group" of the W. Szafer Institute of Botany of the Polish Academy of Sciences, and personally to Prof. Dr Hab. Michał Ronikier for the scientific supervision, technical resources and opportunity to conduct the laboratory part of the research within the framework of a scientific institution.

We also thank the Gesellschaft für wissenschaftliche Datenverarbeitung mbH Göttingen (GWDG) for computational resources (high-performance computing).

Useful comments of the reviewers are greatly appreciated.

SUPPLEMENTARY MATERIAL

This article includes following Supplementary Materials: Matrices S1–S4 (K1–K3, combined); Figures S1–S4 (representing STRUCTURE results) and Haplotype networks S5–S8 (for TCS, Nj-Net, Minimum Spanning Network, Minimum Joining Network, respectively) available as: [ukrbo-tj83-03-161-S1-S8.zip](https://github.com/AndriiTarieiev/Dryas_AFLP). Supplemental data are also available on GitHub by the following link: https://github.com/AndriiTarieiev/Dryas_AFLP

ETHICS DECLARATION

The authors declare no conflict of interest.

ORCID

A. Tarieiev  <https://orcid.org/0000-0002-1123-7746>

R. Cherepanyn  <https://orcid.org/0000-0002-2227-3697>

- Benham J.J. 2001. *Genographer*. Montana State University. Available at: <http://hordeum.oscs.montana.edu/genographer/> (Accessed 18 February 2026).
- Bezusko L.G., Mosyakin S.L., Bezusko A.G. 2011. *Patterns and trends of development of the plant cover of Ukraine in the Late Pleistocene and Holocene*. Kyiv: Alterpress, 448 pp. [Безузько Л.Г., Мосякін С.Л., Безузько А.Г. 2011. *Закономірності та тенденції розвитку рослинного покриву України у пізньому плейстоцені та голоцені*. Київ: Альтерпрес, 448 с.]
- Bezusko L.G., Tsybalyuk Z.M., Nitsenko L.M. 2025. Participation of some rare and relict plant species in the palynofloras of the Allerød–Holocene deposits of the Forest-Steppe zone of Ukraine. *Ukrainian Botanical Journal*, 82(3): 225–233. [Безузько Л.Г., Цимбалюк З.М., Ниценко Л.М. 2025. Участь деяких рідкісних та реліктових видів рослин у палінофлорах відкладів аллереду–голоцену Лісостепової зони України. *Український ботанічний журнал*, 82(3): 225–233.] <https://doi.org/10.15407/ukrbotj82.03.225>
- Chase M.W., Hills H.H. 1991. Silica gel: An ideal material for field preservation of leaf samples for DNA studies. *Taxon*, 40: 215–220. <https://doi.org/10.2307/1222975>
- Chopyk V.I. 1976. *High mountain flora of the Ukrainian Carpathians*. Kyiv: Naukova Dumka, 270 pp. [Чопик В.І. 1976. *Високогірна флора Українських Карпат*. Київ: Наукова думка, 270 с.]
- Cieślak E., Ronikier M., Koch M.A. 2007. Western Ukrainian *Cochlearia* (Brassicaceae): The identity of an isolated edge population. *Taxon*, 56(1): 112–118.
- Cieślak E., Cieślak J., Szelaż Z., Ronikier M. 2015. Genetic structure of *Galium cracoviense* (Rubiaceae): a naturally rare species with an extremely small distribution range. *Conservation Genetics*, 16: 929–938. <https://doi.org/10.1007/s10592-015-0711-7>
- Chhatre V.E., Emerson K. J. 2017. StrAuto: Automation and Parallelization of STRUCTURE Analysis. *BMC Bioinformatics*, 18: 192. <https://doi.org/10.1186/s12859-017-1593-0>
- Cherepanyn R.M. 2017. *Arctic-alpine plant species of the Ukrainian Carpathians*. Ivano-Frankivsk: Publishing House of the Vasyl Stefanyk Precarpathian National University, 92 p. [Черепанин Р.М. *Аркто-альпійські види рослин Українських Карпат*. Івано-Франківськ: Видавництво Прикарпатського національного університету імені Василя Стефаника, 92 с.] <https://doi.org/10.5281/zenodo.7533738>
- Cherepanyn R.M. 2018. Effect of climate changes on the habitat of rare arctic-alpine plant species in the high mountain part of the Ukrainian Carpathians. *Studia Biologica*, 12(1): 73–86. <https://doi.org/10.30970/sbi.1201.544>
- Cherepanyn R.M. 2019. Changes in population vitality of rare arctic-alpine plant species in high mountain part of the Ukrainian Carpathians under influence of climatic factors. *Studia Biologica*, 13(1): 117–128. <https://doi.org/10.30970/sbi.1301.582>
- Clement M., Snell Q., Walke P., Posada D., Crandall K. 2002. TCS: estimating gene genealogies. In: *Proceedings of the 16th International Parallel and Distributed Processing Symposium*. Ft. Lauderdale, FL, USA, pp. 1–7. <https://doi.org/10.1109/IPDPS.2002.1016585>
- Clement M.P., Posada D., Crandall K. 2000. TCS: a computer program to estimate gene genealogies. *Molecular Ecology*, 9: 1657–1659. <https://doi.org/10.1046/j.1365-294x.2000.01020.x>
- De Witte L.C., Armbruster G.F., Gielly L., Taberlet P., Stöcklin J. 2012. AFLP markers reveal high clonal diversity and extreme longevity in four key arctic-alpine species. *Molecular Ecology*, 21(5): 1081–1097. <https://doi.org/10.1111/j.1365-294X.2011.05326.x>
- Green Data Book of Ukraine*. 2009. Ed. Ya.P. Didukh. Kyiv: Alterpress, 448 p. [Зелена книга України. 2009. Ред. Я.П. Дідух. Київ: Альтерпрес, 448с.]
- Erschbamer B., Kiebacher T., Mallaun M., Unterluggauer P. 2009. Short-term signals of climate change along an altitudinal gradient in the South Alps. *Plant Ecology*, 202: 79–89. <https://doi.org/10.1007/s11258-008-9556-1>
- Evanno G., Regnaut S., Goudet J. 2005. Detecting the number of clusters of individuals using the software STRUCTURE: a simulation study. *Molecular Ecology*, 14(8): 2611–20. <https://doi.org/10.1111/j.1365-294X.2005.02553.x>
- French N.Yu.S., Biggs P., Holland B., Fearnhead P., Binney B., Fox A., Grove-White D., Leigh J. W., Miller W., Muellner P., Carter P. 2014. Evolution of *Campylobacter* species in New Zealand. In: Eds S.K. Sheppard, G. Méric. *Campylobacter Ecology and Evolution*. Norfolk, England: Caister Academic Press, pp. 221–240.
- Hammer Ø., Harper D.A.T. 2024. *Paleontological Data Analysis*, 2nd ed. Hoboken, NJ, USA; Chichester, West Sussex, UK: Wiley, xii + 376 pp.
- Hammer Ø., Harper D.A.T., Ryan P.D. 2001. PAST: PAleontological STatistics software package for education and data analysis. *Palaeontologia Electronica*, 4(1): 9. https://palaeo-electronica.org/2001_1/past/past.pdf
- Harald P., Gottfried M., Reiter K., Klettner C., Grabherr G. 2007. Signals of range expansions and contractions of vascular plants in the high Alps: observations (1994–2004) at the GLORIA (Global Observation Research Initiative in Alpine Environments) master site Schrankogel, Tyrol, Austria. *Global Change Biology*, 13: 147–156. <https://doi.org/10.1111/j.1365-2486.2006.01282.x>
- Kobiv Yu. 2018. Trends in population size of rare plant species in the alpine habitats of the Ukrainian Carpathians under climate change. *Diversity*, 10(3): 62. <https://doi.org/10.3390/d10030062>
- Kobiv Yu. 2017. Response of rare alpine plant species to climate change in the Ukrainian Carpathians. *Folia Geobotanica*, 52: 217–226. <https://doi.org/10.1007/s12224-016-9270-z>

- Kopelman N.M., Mayzel J., Jakobsson M., Rosenberg N.A., Mayrose I. 2015. Clumpak: a program for identifying clustering modes and packaging population structure inferences across K. *Molecular Ecology Resources*, 15(5): 1179–91. <https://doi.org/10.1111/1755-0998.12387>.
- Kozłowski G. 2024. Understanding Arctic–Alpine Plants from Ecological and Evolutionary Perspectives. *Plants*, 13(11): 1560. <https://doi.org/10.3390/plants13111560>
- Kreuzer M., Tribsch A., Nyffeler R. 2014. Ecological and genetic differentiation of two subspecies of *Saussurea alpina* in the Western Alps. *Alp Botany*, 124: 49–58. <https://doi.org/10.1007/s00035-014-0128-9>
- Kück P., Greve C., Misof B., Gimnich F. 2012. Automated masking of AFLP markers improves reliability of phylogenetic analyses. *PLoS ONE*, 7(11): e49119. <https://doi.org/10.1371/journal.pone.0049119>
- Kuyak V., Shtupun V. 2021. Transformation processes in alpine phytocoenoses of the Ukrainian Carpathians under reservation and climate changes. *Bulletin of Lviv National University. Biological series*, 85: 59–69. [Кияк В., Штупун В. 2021. Трансформаційні процеси в альпійських фітоценозах Українських Карпат за умов заповідання та кліматичних змін. *Вісник Львівського університету. Серія біологічна*, 85: 59–69.] <https://doi.org/10.30970/vlubs.2021.85.07>
- Leigh J.W., Bryant D. 2015. PopART: Full-feature software for haplotype network construction. *Methods in Ecology and Evolution*, 6(9): 1110–1116. <https://doi.org/10.1111/2041-210X.12410>
- Li Y., Liu J. 2018. STRUCTURESELECTOR: A web-based software to select and visualize the optimal number of clusters using multiple methods. *Molecular Ecology Resources*, 18: 176–177. <https://doi.org/10.1111/1755-0998.12719>
- Liu J., Zhong X. 2024. Population epigenetics: DNA methylation in the plant omics era. *Plant Physiology*, 194(4): 2039–2048. <https://doi.org/10.1093/plphys/kiae089>
- Liu T., Conesa A. 2025. Profiling the epigenome using long-read sequencing. *Nature Genetics*. 57, 27–41. <https://doi.org/10.1038/s41588-024-02038-5>
- Lynch M., Milligan B.G. 1994. Analysis of population genetic structure with RAPD markers. *Molecular Ecology*, 3: 91–99. <https://doi.org/10.1111/j.1365-294X.1994.tb00109.x>
- Marcysiak K. 2010. Arctic-alpine plants and climate change in Europe. *Wiadomości Botaniczne*, 54(3/4): 21–29.
- McGraw J.B., Antonovics J. 1983. Experimental ecology of *Dryas octopetala* ecotypes: II. A demographic model of growth, branching and fecundity. *Journal of Ecology*, 71(3): 899–912. <https://doi.org/10.2307/2259600>
- McGraw J.B. 1987. Experimental ecology of *Dryas octopetala* ecotypes. IV. Fitness response to reciprocal transplanting in ecotypes with differing plasticity. *Oecologia*, 73(3): 465–468.
- Meudt H.M., Clarke, A.C. 2007. Almost forgotten or latest practice? AFLP applications, analyses and advances. *Trends in Plant Science*, 12(3): 106–117. <https://doi.org/10.1016/j.tplants.2007.02.001>
- Mueller U.G., Wolfenbarger L.L. 1999. AFLP genotyping and fingerprinting. *Trends in Ecology & Evolution*, 14: 389–394. [https://doi.org/10.1016/S0169-5347\(99\)01659-6](https://doi.org/10.1016/S0169-5347(99)01659-6)
- Novembre J. 2016. Pritchard, Stephens, and Donnelly on population structure. *Genetics*, 204(2): 391–393. <https://doi.org/10.1534/genetics.116.195164>
- Pauli H., Gottfried M., Reiter K., Klettner C., Grabherr G. 2007. Signals of range expansions and contractions of vascular plants in the high Alps: observations (1994–2004) at the GLORIA master site Schrankogel, Tyrol, Austria. *Global Change Biology*, 13: 147–156. <https://doi.org/10.1111/j.1365-2486.2006.01282.x>
- Paun O., Schönswetter P. 2012. Amplified fragment length polymorphism: an invaluable fingerprinting technique for genomic, transcriptomic, and epigenetic studies. *Methods in Molecular Biology*, 862: 75–87. https://doi.org/10.1007/978-1-61779-609-8_7
- Peakall R., Smouse P.E. 2006. GENALEX 6: genetic analysis in Excel. Population genetic software for teaching and research. *Molecular Ecology Notes*, 6: 288–295. <https://doi.org/10.1111/j.1471-8286.2005.01155.x>
- Peakall R., Smouse P.E. 2012. GenALEX 6.5: genetic analysis in Excel. Population genetic software for teaching and research — an update. *Bioinformatics*, 28: 2537–2539. <https://doi.org/10.1093/bioinformatics/bts460>
- Porrás-Hurtado L., Ruiz Y., Santos C., Phillips C., Carracedo Á., Lareu, M.V. 2013. An overview of STRUCTURE: applications, parameter settings, and supporting software. *Frontiers in Genetics*, 4(98): 1–13. <https://doi.org/10.3389/fgene.2013.00098>
- Pucker B., Irisarri I., de Vries J., Xu B. 2022. Plant genome sequence assembly in the era of long reads: Progress, challenges and future directions. *Quantitative Plant Biology* 3: e5. <https://doi.org/10.1017/qpb.2021.18>
- Puechmaillie S.J. 2016. The program STRUCTURE does not reliably recover the correct population structure when sampling is uneven: subsampling and new estimators alleviate the problem. *Molecular Ecology Resources*, 16: 608–627. <https://doi.org/10.1111/1755-0998.12512>
- Raj A., Stephens M., Pritchard J.K. 2014. fastSTRUCTURE: Variational inference of population structure in large SNP data sets. *Genetics*, 197: 573–589. <https://doi.org/10.1534/genetics.114.164350>
- Reynolds J., Weir B.S., Cockerham C.C. 1983. Estimation of the coancestry coefficient: basis for a short-term genetic distance. *Genetics*, 105(3): 767–779. <https://doi.org/10.1093/genetics/105.3.767>
- Springer J.C., Parfitt B.D. 2014. *Dryas*. In: Flora of North America Editorial Committee (eds.), *Flora of North America north of Mexico*, Vol. 9. New York; Oxford: Oxford University Press, pp. 326–331. http://www.efloras.org/florataxon.aspx?flora_id=1&taxon_id=110971

- Sucher N.J., Hennell J.R., Carles M.C. (eds.) 2012. *Plant DNA fingerprinting and barcoding: Methods and protocols*. [Methods in Molecular Biology series]. New York; Dordrecht; Heidelberg; London: Springer Science+Business Media, LLC, x + 202 pp. <https://doi.org/10.1007/978-1-61779-609-8>
- Tsaryk Y., Kyyak V., Dmytrakh R., Bilonoha V. 2004. Generative reproduction of plant populations of the Carpathian highlands as a sign of their viability. *Bulletin of Lviv National University. Biological series*, 36: 50–56. [Царик Й., Кияк В., Дмитрах Р., Білонога В. 2004. Генеративне розмноження популяцій рослин високогір'я Карпат як ознака їх життєздатності. *Вісник Львів. університету. Серія біологічна*, 36: 50–56.]
- Varsamis G., Merou T., Karapatzak E., Papageorgiou A.C., Fotiadis G., Tsiftsis S. 2021. Genetic diversity of alpine *Dryas octopetala* populations at their southern distribution limit in Europe. *Nordic Journal of Botany*, 2021: e03150. <https://doi.org/10.1111/njb.03150>
- Vekemans X. 2002. AFLP-SURV version 1.0. Distributed by the author. Laboratoire de Génétique et Ecologie Végétale, Université Libre de Bruxelles, Belgium. Available at: https://ebe.ulb.be/ebe/AFLP-SURV_files/manual_AFLPsurv.pdf (Accessed 18 February 2026).
- Vekemans X., Beauwens T., Lemaire M., Roldan-Ruiz I. 2002. Data from amplified fragment length polymorphism (AFLP) markers show indication of size homoplasy and of a relationship between degree of homoplasy and fragment size. *Molecular Ecology*, 11: 139–151. <https://doi.org/10.1046/j.0962-1083.2001.01415.x>
- Vos P., Hogers R., Bleeker M., Reijans M., van de Lee T., Hornes M., Frijters A., Pot J., Peleman J., Kuiper M., Zabeau M. 1995. AFLP: a new technique for DNA fingerprinting. *Nucleic Acids Research*, 23(21): 4407–4414. <https://doi.org/10.1093/nar/23.21.4407>
- White D., Pirro S. 2022. The complete genome sequences of three species of Mountain Avens (*Dryas*, Rosaceae). *Biodiversity Genomes*, 07 November 2022. <https://doi.org/10.56179/001c.40366>.

Генетичне різноманіття та структура популяцій аркто-альпійського реліктового виду *Dryas octopetala* (Rosaceae) в Україні за даними AFLP-аналізу

Андрій ТАРЄЄВ^{1,2}, Роман ЧЕРЕПАНИН^{3,4}

¹ Відділ лісової генетики та селекції лісових дерев, Геттінгенський університет, Бюсгенвег 2, Геттінген 37077, Німеччина

² Інститут біології, геоботаніки та ботанічний сад, Університет Мартіна Лютера Галле-Віттенберг, Ам Кірхтор 1, Галле 06108, Німеччина

³ Міжнародний науково-освітній центр "Обсерваторія", Кафедра біології та екології, Карпатський національний університет імені Василя Стефаника, вул. Шевченка 57, Івано-Франківськ 76018, Україна

⁴ WWF-Україна (Громадська спілка "Всесвітній фонд природи Україна"), вул. Раїси Окіпної 4, офіс 170, Київ 02002, Україна

Реферат. Аркто-альпійські рослинні угруповання містять численні ендемічні та реліктові види. Багато з них є чутливими до змін середовища існування і зазнають впливу кліматичних змін, що робить їхнє дослідження особливо актуальним. У цьому дослідженні зосереджено увагу на циркумполярному аркто-альпійському реліктовому диплоїдному чагарничку *Dryas octopetala* в Україні. Було обстежено та відібрано зразки з усіх чотирьох відомих популяцій. Для аналізу генетичного різноманіття, структури популяцій та інших параметрів використано три набори по 50 різних AFLP-маркерів у кожному. Аналізи STRUCTURE у поєднанні з філогенетичною мережею не виявили суттєвих відмінностей у генетичній структурі між популяціями. Водночас популяція на г. Бребенескул демонструє незначні відмінності порівняно з трьома іншими популяціями дріади в Україні. Географічна ізоляція, різні міграційні шляхи та потенційна можливість схрещування можуть пояснювати отримані результати. Для формування більш чіткої картини необхідні додаткові дослідження із застосуванням різних методів (зокрема SSR, повногеномного аналізу SNP, GWAS, WGS).

Ключові слова: *Dryas octopetala*, AFLP, STRUCTURE, PCoA, Карпати, популяції, Україна